# Antibacterial Effect of Iranian Green Tea and Black Tea against Actinobacillus actinomycetemcomitans, Porphyromonas gingivalis and Prevotella intermedia

Mohammad Niakan (PhD) Department of Microbiology, Faculty of Medicine, Shahed University, Tehran, Iran

Noushin Jalayer Naderi (DMD) Department of Oral and Maxillofacial Pathology, Faculty of Dentistry, Shahed University, Tehran, Iran

Hadise Jamshidian (DDS) Faculty of Dentistry, Shahed University, Tehran, Iran

Fateme Jafariazad (MSc) Department of Microbiology, Faculty of Medicine, Shahed University, Tehran, Iran

**Corresponding author:** Noushin Jalayer Naderi

Email: jalayer@shahed.ac.ir

Tel: +989122148468

Address: No.39, Department of Oral and Maxillofacial Pathology, Faculty of Dentistry, Shahed University, Italia Street, Tehran, Iran

**Received :** 01 May 2016 **Revised:** 01 Sep 2016 **Accepted:** 13 Sep 2016

#### ABSTRACT

**Background and Objective:** Anaerobic bacteria are the main cause of periodontitis. It has been shown that green tea and black tea have antibacterial effect. The aim of this study was to determine he antibacterial effect of Iranian green tea and black tea against *Actinobacillus actinomycetemcomitans, Porphyromonas gingivalis* and *Prevotella intermedia.* 

**Methods:** Aqueous and methanolic extracts of Iranian green tea and black tea at concentrations ranging from 10 to 500 mg/ml were tested against standard strains of *A. actinomycetemcomitans* (ATCC 33384), *P. gingivalis* (ATCC 33227) and *P. intermedia* (ATCC 25671) using agar disk diffusion, broth microdilution and determination of minimum inhibitory concentration.

**Results:** *P. gingivalis, A. actinomycetemcomitans* and *P. intermedia* were sensitive to the methanolic extract of Iranian green tea at concentrations of 100-500 mg/ml, 10-500 mg/ml and 50-500mg/ml, respectively. *P. gingivalis, A. actinomycetemcomitans* and *P. intermedia* were sensitive to the methanolic extract of Iranian black tea at concentrations of 200-500 mg/ml, 20-500 mg/ml and 200-500 mg/ml, respectively. In addition, *P. gingivalis, A. actinomycetemcomitans* and *P. intermedia* were sensitive to the aquatic extract of Iranian green tea at concentrations of 200-500 mg/ml, negretively. In addition, *P. gingivalis, A. actinomycetemcomitans* and *P. intermedia* were sensitive to the aquatic extract of Iranian green tea at concentrations of 200-500mg/ml, 100-500 mg/ml and 200-500 mg/ml, respectively.

**Conclusion:** The aquatic and alcoholic extracts of Iranian green tea and black tea have antibacterial activity against *A. actinomycetemcomitans*, *P. intermedia* and *P. gingivalis*. Therefore, incorporation of Iranian black tea as an effective native herb could be beneficial for prevention of oral cavity diseases.

Keywords: Tea, Green Tea, Antibacterial Agents, Anaerobic Bacteria.

### INTRODUCTION

Periodontitis is an inflammatory disease that could lead to destruction of supporting structures of teeth. Anaerobic bacteria are the main cause of initiation and progression of (1-3).Actinobacillus periodontitis actinomycetemcomitans, *Porphyromonas* gingivalis and Prevotella intermedia are among the main anaerobic pathogens involved in periodontitis development (4). Prevention and treatment of periodontitis has been a challenging issue in modern dentistry. While mouthwashes are useful tools for controlling the oral microflora in periodontitis, they have some disadvantages including tooth discoloration and unpleasant taste. Natural plants with therapeutic properties could be considered as alternatives for such products. These plants are easily available and cost effective.

Tea (*Camellia sinensis*) is the most popular drink in Iran. In general, black tea is fully fermented, oolong is partially fermented, and green tea is not fermented or only minimally fermented (5-6).

It has been shown that all types of tea have antibacterial. antifungal and antioxidant properties (7-9). Tea-derived catechins and epigallocatechin gallate (EGCG) can inhibit the activity of periodontal pathogens. In addition, tea polyphenols have a positive effect on the inflammatory response in periodontal structures (10-12). It has been shown that growth conditions including soil composition, temperature and cultivating methods can affect the chemical properties of tea (13). Some studies have previously demonstrated the antibacterial effect of aquatic and alcoholic extracts of Iranian green tea on periodontopathic bacteria (14-15). However, there is limited information about the antibacterial effect of Iranian black tea. Black tea is a popular beverage in Iran, especially among the older population. The aim of this study was to determine the antibacterial effect of Iranian green tea and black tea on a number of periodontitis-causing anaerobic bacteria.

# MATERIAL AND METHODS

First, tea leaves were collected from Baz-Kia-Gurab region in Gillan Province, Iran. The samples were verified by the Department of Pharmacognosy (School of Pharmacy) at Shahid Beheshti University of Medical Sciences, Iran. For alcoholic extraction of green tea and black tea, 500 ml of 70% methanol (Merck Co. Germany) were added to a sterile flask containing 100 g of chopped, powdered green tea and black tea samples, separately. The mixture was place at room temperature for two days. After filtering the mixture through No.1 filter papers (Whatman Co. Germany), the extract was dried in water bath (GFL Co., Germany) at 70 °C. The dried powder obtained was kept at 4 °C in sealed vials for future analysis. The following concentrations of the extracts were prepared from the stock solution: 1% (10mg/ml), 2% (20mg/ml), 5% (50mg/ml), 10% (100mg/ml), 20% (200mg/ml), 30% (300mg/ml), 40% (400mg/ml) and 50% (500mg/ml) (16).

Aqueous extraction of green tea and black tea was done separately. First, 1L of sterilized boiling water was added to 100 g of tea. After 4 hours, the mixture was filtered by No.1 filter paper (Whatman Co., Germany), and then dried in water bath (GFL Co., Germany) at 70 °C. A range of different concentrations was prepared from the stock solution similar to the previous step.

Standard strains of A. actinomycetemcomitans (ATCC 33384), P. gingivalis (ATCC 33227) and P. intermedia (ATCC 25671) were obtained from bacterial collection of School of Medicine, Shahed University, Iran. The bacteria were inoculated into solid and aqueous media containing 41 g/L Brucella agar, 52g/L BHA, 44 g/L anaerobic blood agar, 30 g/L thioglycollate fluid, 29 g/L thioglycollate broth and 30 g/L trypticase soy broth (Merck Co. Germany). The samples were inoculated into 5 mg/ml hemincontaining yeast extract (Sigma Co. Germany), and kept at 37°C in anaerobic conditions by Gas-pak (Merck Co. Germany). Then, 10 µg of vitamin K and 100 ml defibrinated sheep blood (Bahar afshan Co. Iran) were added to the anaerobic blood agar (Merck Co. Germany).

All media were kept under anaerobic condition at 4 °C for future testing. The antimicrobial activity of the alcoholic and aqueous extracts of green tea and black tea was evaluated using disk diffusion, well diffusion and minimum inhibitory concentrations (MICs). The antibacterial effect of green tea and black tea was evaluated against standard strains of *A. actinomycetemcomitans* (ATCC 33384), *P.*  gingivalis (ATCC 33227) and P. intermedia (ATCC 25671). Sterile blank paper discs were soaked in the extracts. Then, 20 µl of 0.5 McFarland bacterial suspension was spread on the surface of sterile Muller Hinton agar plates (Merck Co. Germany). Paper disks (Padtan Teb Co. Iran) containing different concentrations of the extracts were placed on the surface of each plate. The plates were incubated at 37 °C for 72 hours in anaerobic condition. The antibacterial activity of the extracts against the bacteria was assessed by measuring the growth inhibition zone around the disks. MIC of crude extracts was determined by broth dilution method. Briefly, 1ml of sterile trypticase soy broth (Merck Co. Germany) was transferred to a sterile test tube. Then, 10 µl of 0.5 McFarland bacterial suspension was inoculated into a test tube containing 1ml of sterile trypticase soy broth Co. Germany). Different (Merck concentrations of the extracts were added to each test tube. The content of the tubes was mixed thoroughly by gentle shaking. The test tubes were incubated at 37 °C for 3-7 days in anaerobic condition. A tube without bacteria was also prepared as negative control. Bacterial cultures were swabbed on Mueller Hinton agar plates (Merck Co. Germany) by diffusion method. disk Moreover, the antibacterial activity of the extracts was that vancomycin, compared with of clindamycin, ciprofloxacin, ampicillin, tetracycline, amoxicillin, gentamycin, kanamycin and penicillin paper disks (Mast Co., UK).

# RESULTS

As shown in Table 1, *P. gingivalis, A. actinomycetemcomitans* and *P. intermedia* were sensitive to the methanolic extract of

green tea at concentrations of 100-500 mg/ml, 10-500 mg/ml and 50-500 mg/ml, respectively. The MIC of methanolic extract of green tea for Р. gingivalis, Α. actinomycetemcomitans and P. intermedia was 50 mg/ml, 20 mg/ml and 10 mg/ml, respectively (Table 2). P. gingivalis, A. actinomycetemcomitans and P. intermedia were sensitive to the aquatic extract of Iranian green tea at concentration of 200-500 mg/ml, 100-500 mg/ml and 200-500 mg/ml. respectively (Table 1). The MIC of the aquatic extract of green tea for P. gingivalis, A. actinomycetemcomitans and P. intermedia was 50 mg/ml, 50 mg/ml and 20 mg/ml, respectively (Table 2).

P. gingivalis, A. actinomycetemcomitans and *P. intermedia* were sensitive to the methanolic extract of Iranian black tea at concentrations of 200-500 mg/ml, 20-500 mg/ml and 50-500 mg/ml, respectively (Table 1). The MIC of the methanolic extract of black tea for P. gingivalis, A. actinomycetemcomitans and P. intermedia was 100 mg/ml, 50 mg/ml and 20 mg/ml, respectively (Table 2). As shown in gingivalis, Table 1. Ρ. Α. actinomycetemcomitans and P. intermedia were sensitive to the aquatic extract of black tea at concentrations of 200-500 mg/ml, 100-500 mg/ml and 200-500 mg/ml, respectively. The MIC of the aquatic extract of black tea for P. gingivalis, A. actinomycetemcomitans and P. intermedia was 300 mg/ml, 20 mg/ml and 100 mg/ml, respectively (Table 2).

In addition, the methanolic and aquatic extracts of green tea had more antibacterial effect at lower concentrations compared with black tea (Table 2). Moreover, comparison of the antibiotic activity of green tea and black tea with antibiotics is shown in Table 3.

		10	20	50	100	200	300	400	500
		mg/ml							
Methanolic extract of green tea	Pg	-	-	-	7mm	9mm	12mm	14mm	15mm
	Aa	9mm	10mm	12mm	13mm	13mm	14mm	14mm	15mm
	Pi	-	-	8mm	10mm	10mm	12mm	13mm	14mm
Aqueous extract of green tea	Pg	-	-	-	-	8mm	8mm	10mm	12mm
	Aa	-	-	-	8mm	9mm	10mm	11mm	14mm
	Pi	-	-	-	-	8mm	8mm	12mm	12mm
Methanolic extract of black tea	Pg	-	-	-	-	8mm	10mm	12mm	15mm
	Aa	-	10mm	12mm	12mm	11mm	12mm	12mm	16mm
	Pi	-	-	-	-	10mm	11mm	12mm	14mm
Aqueous extract of black tea	Pg	-	-	-	-	7mm	7mm	8mm	10mm
•	Aa	-	-	-	7mm	7mm	8mm	9mm	10mm
	Pi	-	-	-	-	8mm	9mm	10mm	12mm

Table 1- Antimicrobial activity of methanolic and aquatic extracts of Iranian green tea and black tea by well diffusion method

Pg: P. gingivalis

Aa: A. actinomycetemcomitan

Pi: P. intermedia

		10	20	50	100	200	300	400	500
		mg/ml							
Methanolic extract of green tea	Pg	+	+	-	-	-	-	-	-
_	Aa	-	-	-	-	-	-	-	-
	Pi	-	-	-	-	-	-	-	-
Aqueous extract of green tea	Pg	+	+	-	-	-	-	-	-
	Aa	+	+	-	-	-	-	-	-
	Pi	+	-	-	-	-	-	-	-
Methanolic extract of black tea	Pg	+	+	+	-	-	-	-	-
	Aa	+	+	-	-	-	-	-	-
	Pi	+	+	-	-	-	-	-	-
Aqueous extract of black tea	Pg	+	+	+	+	+	-	-	-
	Aa	+	-	-	-	-	-	-	-
	Pi	+	+	+	-	-	-	-	-

Table 2- MIC of methanolic and aqueous extracts of Iranian green tea and black tea against the bacteria tested

Pg: *P. gingivalis* 

Aa: A. actinomycetemcomitan

Pi: P. intermedia

Table 3- Comparison of antimicrobial activity of antibiotic discs and the extracts against the bacteria tested by disk diffusion method

	AMX	AM	V30	TE	CP	K30	GM 10mg	CC	Р
	25mg	10mg	mg	30mg	5mg	mg	_	2mg	10mg
P. gingivalis	20mm	16mm	16mm	14mm	17mm	-	-	-	-
A. actinomycetem	28mm	16mm	21mm	18mm	30mm	10mm	10mm	-	-
comitans									
P. intermedia	18mm	15mm	15mm	14mm	23mm	10mm	20mm	12mm	-
MV. Amoviaillin AM. A	mniaillin X	V. Voncomvoi	n TF						

AMX: Amoxicillin, AM: Ampicillin, V: Vancomycin, TE

: Tetracycline, CP: Ciprofloxacin, K: Kanamycin, GM:

Gentamycin, CC: Clindamycin, P: Penicillin

#### DISCUSSION

In present study, we examined the antibacterial activity of the methanolic and aquatic extracts of Iranian green tea and black tea against a number of anaerobic periodontal pathogens. The results showed that P. gingivalis, A. actinomycetemcomitans and P. intermedia are sensitive to the extracts, with more sensitivity observed against the extracts of Iranian green tea. These findings are in agreement with previous reports on the aquatic extract of green tea (14-17). Highest antibacterial activity by both extracts of Iranian green tea and black tea was observed at concentration of 200 mg/ml, which is in agreement with results of Sakanaka et al. (18). The MIC of aquatic extracts of Iranian green tea and black tea was 50 mg/ml and 100 mg/ml, respectively. However, Araghizadeh et al. found that gingivalis, Р. Α. actinomycetemcomitans and P. intermedia are most sensitive to the aquatic extract of green tea at concentration of 12.5-50 mg/ml (14). The MIC of aquatic extract of green tea for *P*. gingivalis, A. actinomycetemcomitans and P. intermedia was 50 mg/ml, 50 mg/ml and 20 mg/ml, respectively. These findings are inconsistent with findings of Araghizadeh et al. (14). This could be due to the differences in the methods used. For instance, we have used bacterial stocks, while Araghizadeh et al.

examined the aquatic extract of green tea on oral-derived bacteria.

According to previous studies, consumption of green tea reduces the periodontal destruction and significantly improves periodontal indices (15 -19). The antibacterial activity of tea is mainly attributed to its chemical components such as polyphenolic compounds and catechin (9, 20-22). It has been shown that the bactericidal activity of green tea could also be due to the impact of EGCG on bacterial membrane (23).

*P. gingivalis* is a key microorganism involved in initiation of periodontitis. The bacterium produces proteolytic enzymes such as collagenases and dipeptidyl aminopeptidase IV. Tea-derived polyphenols inhibit proteases production in *P. gingivalis* (24). Sakanaka et al. demonstrated the effect of green teaderived polyphenols, especially EGCG on the growth and adherence of *P. gingivalis* onto the oral buccal cells (18). In addition, green teaderived catechin has bactericidal effects on *Prevotella spp.* with MIC of 1.0 mg/ml (25).

Okamoto et al. suggested that the inhibitory effect of green tea on the activity of *P*. *intermedia* might be due to the impact of EGCG on tyrosine phosphatase (26). The catechin content of green tea is more than that of black tea and oolong tea. This difference

originates from oxidation of tea leaves during fermentation. The fermentation process leads to conversion of flavanoids into aflevine and arubigin (6).

Various methods and therapeutic agents have been used for eliminating periodontopathic bacteria in mouth. However, they all have some disadvantages such as tooth discoloration. Herbal-based mouthwashes could have some beneficial effects for eradication of the periodontopathic microflora. Such products are safer and more costeffective compared with currently available chemical counterparts. Based on the results of our study and its antibacterial properties, tea could be incorporated into mouthwashes for prevention or even treatment of oral diseases. However, in vivo studies should be performed on the effect of different concentrations of green tea and black tea extracts on P. gingivalis, A. actinomycetemcomitans and P. intermedia to verify the results of the present study.

#### REFERENCES

1. Haffajee AD, Socransky SS, Taubman MA, Sioson J, Smith DJ. *Patterns of antibody response in subjects with periodontitis*. Oral Microbiol Immunol. 1995; 10(3): 129-37.

2. Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent RL Jr. *Microbial complexes in subgingival plaque*. J Clin Periodontol. 1998; 25(2):134-44.

3. Chapple IL. *Reactive oxygen species and antioxidants in inflammatory diseases*. J Clin Periodontol. 1997; 24(5): 287-96.

4. Patil KG, Metgud SC. *Effects of Areca Nut Extracts on Phagocytosis of Actino bacillus actionmycetem comitans ATCC 33384 by Neutrophils in Patients with Chronic Periondontitis.* J Clin Diagn Res. 2013; 7(10): 2153-6. doi: 10.7860/JCDR/2013/5694.3456.

5. Chacko SM, Thambi PT, Kuttan R, Nishigaki I. *Beneficial effects of green tea: a literature review.* Chin Med 2010; 5: 13. doi: 10.1186/1749-8546-5-13.

6. Cabrera C, Artacho R, Gimenez R. *Beneficial effects of green tea–a review*. J Am Coll Nutr. 2006; 25(2): 79-99.

7. Schneider C, Segre T. *Green tea: potential health benefits.* Am Fam Physician. 2009; 79(7): 591-4.

8. Stoicov C, Saffari R, Houghton J. *Green tea inhibits Helicobacter growth in vivo and in vitro*. Int J Antimicrob Agents. 2009; 33(5): 473-8. doi: 10.1016/j.ijantimicag.2008.10.032.

9. Naderi NJ, Niakan M, KharaziFard MJ, Zardi S. Antibacterial activity of Iranian green and black tea on streptococcus mutans: an in vitro study. J Dent (Tehran). 2011; 8(2): 55-9.

# CONCLUSION

The methanolic and aquatic extracts of Iranian green tea and black tea have antibacterial effect on *P. gingivalis, A. actinomycetemcomitans* and *P. intermedia* in a dose-dependent manner. Iranian black tea can be used as an effective herbal mouthwash to prevent oral cavity diseases.

# ACKNOWLEDGMENTS

The study has been supported by the Deputy of Research at Shahed University (Grant No.118). The authors would like to thank Dr Eshraghi (Tehran University of Medical Sciences) and the Department of Microbiology at Shahed University for their kind assistance.

# **CONFLICT OF INTEREST**

The authors declare that there is no conflict of interest.

10. Yun JH, Pang EK, Kim CS, Yoo YJ, Cho KS, Chai JK, et al. *Inhibitory effects of green tea polyphenol* (-)-*epigallocatechin gallate on the expression of matrix metalloproteinase-9 and on the formation of osteoclasts.* J Periodontal Res. 2004; 39(5): 300-7.

11. Yun JH, Kim CS, Cho KS, Chai JK, Kim CK, Choi SH. (-)-*Epigallocatechin gallate induces apoptosis, via caspase activation, in osteoclasts differentiated from RAW 264.7 cells.* J Periodontal Res. 2007; 42(3): 212-8.

12. Krahwinkel T, Willershausen B. *The effect of sugar-free green tea chew candies on the degree of inflammation of the gingiva*. Eur J Med Res. 2000; 5(11): 463-7.

13. Khokhar S, Magnusdottir SGM. Total phenol, catechin, and caffeine contents of teas commonly consumed in the United Kingdom. J Agric Food Chem. 2002; 50: 565-570.

14. Araghizadeh A, Kohanteb J, Fani MM. *Inhibitory* activity of green tea (Camellia sinensis) extract on some clinically isolated cariogenic and periodontopathic bacteria. Med Princ Pract. 2013; 22(4): 368-72.

 Jenabian N, Moghadamnia AA, Karami E, Mir A PB. The effect of Camellia Sinensis (green tea) mouthwash on plaque-induced gingivitis: a single-blinded randomized controlled clinical trial. Daru. 2012; 20(1): 39.

16. CLSI. Performance Standards for Antimicrobial Susceptibility Testing; Twenty -second informational supplement. CLSI document M100-S22. Wayne, PA, USA: Clinical and Laboratory Standards Institute. 2012; 122-124.

17. Sakanaka S, Okada Y. Inhibitory effects of green tea polyphenols on the production of a virulence factor of the periodontal-disease-causing anaerobic bacterium Porphyromonas gingivalis. J Agric Food Chem. 2004; 52(6):1688-92.

18. Sakanaka S, Aizawa M, Kim M, Yamamoto T. Inhibitory effects of green tea polyphenols on growth and cellular adherence of an oral bacterium, Porphyromonas gingivalis. Biosci Biotechnol Biochem. 1996; 60 (5): 745-9.

19. Kushiyama M , Shimazaki Y, Murakami M, Yamashita Y. *Relationship between intake of green tea and periodontal disease*. J Periodontol. 2009; 80(3): 372-7.

20. Takabayashi F, Harada N, Yamada M, Murohisa B, Oguni I. *Inhibitory effect of green tea catechins in combination with sucralfate on Helicobacter pylori infection in Mongolian gerbils.* J Gastroenterol. 2004; 39(1): 61-3.

21. Taylor PW, Hamilton-Miller JM, Stapleton PD. *Antimicrobial properties of green tea catechins*. Food Sci Technol Bull. 2005; 2: 71-81.

22. Yoshino K, Nakamura Y, Ikeya H, Sei T, Inoue A, Sano M, Tomita I. *Antimicrobial activity of tea extracts on cariogenic bacterium (Streptococcus mutans).* J Food Hyg Soc Japan. 1996; 37(2): 104-108.

23. Ikigai H, Nakae T, Hara Y, Shimamura T. *Bactericidal catechins damage the lipid bilayer*. Biochim Biophys Acta. 1993; 1147(1): 132-6.

24. Grenier D, La VD. Proteases of Porphyromonas gingivalis as important virulence factors in periodontal disease and potential targets for plant-derived compounds: a review article. Curr Drug Targets. 2011; 12(3): 322-31.

25. Hirasawa M, Takada K, Makimura M, OtakeS. *Improvement of periodontal status by green tea catechin using a local delivery system: A clinical pilot study.* J Periodontal Res. 2002; 37(6): 433-8.

26. Okamoto M, Leung KP, Ansai T, Sugimoto A, Maeda N. *Inhibitory effects of green tea catechins on protein tyrosine phosphatase in Prevotella intermedia*. Oral Microbiol Immunol. 2003; 18(3): 192-5.