# A-10-702-3 Original article

The effect of running and starvation intervention on atherogenic index and xbp1 gene change in liver endoplasmic reticulum of non-alcoholic fatty liver rats.

**Running title:** Regulation of endoplasmic reticulum stress-induced apoptotic pathway activity by running and starvation intervention

#### Mahshad Paziraee

Department of Physical Education and sports sciences, Aliabad katoul Branch, Islamic Azad University, Aliabad Katoul, Iran. mahshad.pazirayi@gmail.com; ORCID: 0009-0003-2993-7286

## \*Habib Asgharpour

Department of Physical Education and sports sciences, Aliabad katoul Branch, Islamic Azad University, Aliabad Katoul, Iran. Habib.asgharpour@gmail.com; ORCID: 0000-0002-4116-981x

#### \*Asra Askari

Department of Physical Education and sports sciences, Gorgan Branch, Islamic Azad University, Gorgan, Iran, askari.asra@gmail.com; ORCID: 0000-0002-4948-9658

## Reza Rezaei Shirazi

Department of Physical Education and sports sciences, Aliabad katoul Branch, Islamic Azad University, Aliabad Katoul, Iran, rezaii725@yahoo.com; ORCID: 0000-0002-0657-2865

## Neda Aghaei Bahman Beglo

Department of Physical Education and sports sciences, Aliabad katoul Branch, Islamic Azad University, Aliabad Katoul, Iran, nedaaghaei@gmail.com; ORCID: 0000-0003-0972-1288

## **Corresponding authors:**

**1.**Dr Habib Asgharpour

Address: Department of Physical Education and sports sciences, Aliabad katoul Branch, Islamic Azad

University, Aliabad Katoul, Iran

Email: Habib.Asgharpour@gmail.com

2.Dr Asra Askari

Address: Department of Physical Education and sports sciences, Gorgan Branch, Islamic Azad University, Gorgan,Iran

Email:askari.asra@gmail.com

Tel: +989111566028

## **Abstract**

**Background**: Running and starvation can have a positive effect on the reticulophagy of the patient's liver tissue. The purpose of this research is the effect of running and starvation intervention on atherogenic index and xbp1 gene change in liver endoplasmic reticulum of non-alcoholic fatty liver rats (NAFLD).

**Methods:** 30 obese male Wistar rats aged 18-20 weeks with an average body weight of  $348 \pm 25.53$  after one week familiarization with the laboratory environment were randomly divided into 6 groups of 5: 1) starvation, 2 and 3) (3 and 5 days of training), 4 and 5) (3 and 5 days of training with starvation) and 6) the control group were divided. All fatty model animals had free access to water and standard pellets (10 gr of food per 100 g of mouse body weight). The statistical test of one-way analysis of variance was used at a significance level of less than 0.05 and the LSD post hoc test was used among the research groups.

**Results:** According to the experimental results and statistical analysis of one-way analysis of variance it showed a significant decrease in the ratio of lipoproteins (VLDL/HDL, LDL/HDL) in all experimental groups compared to the control group, and also a significant decrease in the expression of XBP1 and CHOP genes was observed in the groups of 3 and 5 days of exercise alone and with starvation.

**Conclusion**: Regular exercise for 3 and 5 days per week with starvation can possibly help by reducing the possible activity of genes involved in increasing endoplasmic reticulum stress in NAFLD patients.

Keywords::CHOP; Exercise; Endoplasmic Reticulum; Starvation; Xbp1

## **Introduction:**

The liver is a complex organ that performs many physiological functions (1), including the synthesis, oxidation, and transport of free fatty acids (FFA), triglycerides (TG), cholesterol, and bile acids (BA). It plays a key role in lipid homeostasis (2). These processes act through pathways that lead to oxidative stress, chronic inflammation, and insulin resistance (3).

The reported prevalence of non-alcoholic fatty liver (NAFLD) in Western countries is between 30 and 46%. This disease has also spread in eastern countries and has become one of the public health concerns in these regions (4). NAFLD includes a spectrum of liver damage from steatosis to nonalcoholic steatohepatitis (NASH), which can lead to fibrosis (5). People with NAFLD are also at increased risk of cardiovascular disease, type 2 diabetes, and obesity-related mortality. The exact mechanisms of NAFLD are still not well understood (6). The "multiple hit hypothesis" is currently the most recognized theory to explain the development and progression of the disease. The initial shock leads to simple steatosis, while the subsequent shocks include mitochondrial dysfunction, oxidative stress, adipocytokine changes, lipid peroxidation, Kupffer cell activation, etc., leading to liver cell inflammation and apoptosis, which in Finally, it leads to simple steatosis (7). Recently, and based on accumulated data, it has been shown that disruption of endoplasmic reticulum (ER) homeostasis, or ER stress, is involved in both the development of Steatosis and the progression to NASH (8). ER is a membrane-bound organelle that provides a specialized environment for the production and post-translational modification of secretory and membrane proteins, lipid biosynthesis, and intracellular Ca2+ homeostasis (9). Some physiological and pathological conditions, including temperature and pH changes, accumulation Damaged DNA can cause ER stress (10). ER stress can be divided into three types, including the unfolded protein response (UPR), ER overload response, and sterol regulatory elements along with regulatory responses with protein mediator; ER stress is commonly referred to as the UPR and occurs when folded or unfolded proteins in the ER increase and activate a stress signal that is transmitted through the ER membrane to the nucleus (11). Findings show that membrane receptors ERs recognize the onset of ER stress and initiate the UPR to restore normal ER function. If the stress is prolonged, or the adaptive response fails, apoptotic cell death occurs (12).

As a result of ER stress, cells mainly develop two responses: one leads to cell survival and the other leads to apoptosis (13). Using the survival pathway, cells overcome such adverse effects and maintain homeostasis through the UPR, from They inhibit mRNA transcription, increase the folding capacity of ER and Endoplasmic-reticulum-associated protein degradation (ERAD) to restore homeostasis (14). Under chronic or severe ER stress, the normal functions of ER are not recovered, resulting in cell dysfunction and apoptosis (14) Therefore, the ER is considered a quality control checkpoint and only correctly folded proteins can exit the ER and pass through the secretory pathway. Therefore, any event such as starvation and excessive protein synthesis, accumulation of mutant proteins, depletion of ER calcium, or changes in the redox state that disrupt the folding capacity of the ER triggers a physiological response called the unfolded protein response (UPR). These homeostatic responses cause the production of additional chaperones to increase the folding capacity of ER, increase protein degradation related to the endoplasmic reticulum, and by changing the translation and synthesis of new proteins, reducing protein entry and thus organelle and cell balance (15). Studies show that silencing C/EBP homologous protein (CHOP) reduces liver apoptosis in alcohol-induced diseases and cholestasis-induced fibrosis (16).

CHOP can also regulate the expression of autophagy-related genes in the later stages of starvation, CHOP can prevent the occurrence of autophagy and soon initiate apoptosis (17), however, the role of CHOP in NAFLD is debatable (18). One study found that CHOP can prevent it (19,20). Also, experiments on mice with CHOP show that CHOP is associated with many diseases that cause ER stress (21).

Chronic ER stress interferes with body metabolism by activating lipogenesis and increasing VLDL (22). Research is ongoing to provide alternative non-pharmacological pathways to reduce the risks of NAFDL. Exercise is one of the ways to replace drugs in this disease for any disease, Hutton et al. (23,24) also, Bucky et al. (25) suggested that both aerobic and resistance exercises have similar effects on liver TG in patients with NAFLD. These studies show that different types of exercise help reduce NAFLD. Also, interventional studies have shown that regular exercise can reverse ER disorders (26) and UPR activation has been shown to reduce ER stress (27,28). The UPR is an important mechanism for modulating fatty acid oxidation and lipogenesis (29). Furthermore, chronic fasting conditions in mice have been shown to activate the UPR to regulate lipid metabolism (30). Studies have shown that XBP1 regulates genes involved in various cellular processes, such as ER stress response, secretory function, lipid metabolism, glucose homeostasis, and inflammation (31,32). XBP1 regulates the expression of genes involved in fatty acid synthesis and increases hepatic lipogenesis (33). Several studies have shown that XBP1 plays an important role in adipocyte differentiation by regulating morphological and functional changes during adipogenesis (34). The importance of XBP1s in lipid biosynthesis has been demonstrated. It causes triglyceride (TG) biosynthesis and abnormal fat accumulation (35). Chronic starvation in mice has been shown to activate the UPR to regulate lipid metabolism (36). Also, studies show that exercise up-regulates hepatic XBP1 and SREBP through ERS signaling, thereby reducing lipid accumulation in NAFLD liver (37). The contradictions from human and animal experiments led us to investigate the effect of aerobic running on a treadmill and 4-week starvation in regulating the activity of the apoptotic pathway caused by endoplasmic reticulum stress in the liver of male rats with non-alcoholic fatty liver disease.

#### **Methods**

30 obese male Wistar rats aged 18-20 weeks with an average body weight of  $348 \pm 25.53$  after one week familiarization with the laboratory environment were randomly divided into 6 groups of five, 1: starvation group, 2 and 3: 3 and 5 training days respectively), 4 and 5: (groups 3 and 5 training days with starvation) and 6: control group were divided

All fatty model animals had free access to water and standard pellet food (10 g of food per 100 g of mouse body weight). All maintenance and sacrifice procedures were carried out in the Animal Science Laboratory of Gorgan Medical Sciences. The fasting protocol was applied for one month and every day for 14 hours in the waking cycle (5.5 pm to 5.7 am). In order to induce hunger, the rats in the starvation group were given the same amount of food (10 grams per 100 grams of mouse weight). They received the same food over 10 hours as the other groups received over 24 hours. The entire training course includes two stages of familiarization and main training. For this purpose, perform the test conditions for 15 minutes for a week and exercise for 45-60 minutes on the treadmill, 3 and 5 days a week for 4 weeks. The training of the rats started on a treadmill with

a 0-degree incline. With a speed of 14 meters and after finishing the training sessions, the speed of the treadmill with zero incline reached 16 and 18 meters per minute (38).

Biochemical factors of HDL were measured by enzymatic method and LDL and VLDL by calorimetric method, respectively, using the biochemical kits of Darman Kav and Far Samad manufactured in Iran and BS480 Auto analyzer.

Finally, the ratios of VLDL/HDL and LDL/HDL were calculated for statistical analysis. For molecular investigations at the level of gene expression, RNA was first extracted from the tissues in all investigated groups, according to the protocol of Yekta azma Equipment manufacturer (cat.No:FABRK001 lot.No:K812320822). Then, we measured the quality and quantity of RNA with the Nanodrop device of Golestan University of Medical Sciences and analyzed it with the cDNA synthesis assay kit of Pars Tous Company of Mashhad (parstous.lot:2156, REF: A101161) and then the synthesized cDNA was used to perform the reverse transcription reaction. Was used Expression levels of xbp1 chop genes were measured using real-time steps quantitative method. The primers were made by SYBER Green qPCR master mix (cat.No: YT2552, lot.P2003), and the primers were ordered by Pishgam Biotech company. The glyceraldehyde-3-phosphate dehydrogenase gene (GAPDH) was used as a control gene and the expression level of the desired gene was calculated with the formula  $2-\Delta\Delta CT$  in the following way. First, the threshold cycle of the desired gene of each sample was calculated from the threshold cycle of the homeostasis gene. The sample was subtracted. (ΔCt=Ct Target-Ct Housekeeping) In the next step, the number obtained from delta Ct of each sample was subtracted from the samples to which it needed to be compared (ΔΔCt=ΔCt Target-ΔCt Reference) and we multiply the negative of the obtained number to the power of two Target gene/Reference gene ratio =  $2 - \Delta \Delta CT$  and

We obtain the relative expression of xbp1 chop gene .The primers used are reported in Table 1below. The size of the genes is as follows C/EBP homologous protein gene ID: DDIT3 gene length 150 bp and X-box binding protein 1 gene ID: XBP1 gene length: 601 bp.

Table 1: Sequence of primers used

Genes	Primer sequence $(5' \rightarrow 3')$	Number of nucleotides	Amplicon Size(pb)
Chop-F	GAAAGCAGAAACCGGTCCAAT	21	150
Chop-R	GGATGAGATATAGGTGCCCCC	21	-
XBP1-F	AAACAGAGTAGCAGCGCAGACTGC	24	601
XBP1-R	GGATCTCTAAAACTAGAGGCTTGGTG	26	-
GAPDH-F	CACTGAGCATCTCCCTC ACAA	22	-
GAPDH-R	TGGTATTCGAGAGA AGGGAGG	22	-

In order to check the descriptive statistics of the mean and standard deviation and estimate the inferential statistics from the one-way analysis of variance (p) and the LSD follow-up test among the research groups, it was shown in Tables 3 and 4, respectively.

#### **Result:**

The results obtained from Table (2) and the average of Graph 1(A,B) showed the lowest mean LDL/HDL ratio in the 5-day training plus fasting group (0.28±0.61) and the lowest VLDL/HDL value was also shown in the 5-day training plus fasting group (0.12±0.286), which indicates the better effects of combined training and fasting. Also, the lowest mean XBP1 gene expression (0.13±0.20) was seen in the fasting plus 5-day training group and a greater decrease in the mean chaperone gene expression (0.06±0.15) was seen in the 5-day training plus fasting group. Genes involved in the inflammatory pathway promote autophagy in non-alcoholic fatty liver patients with exercise and fasting.

The results of one-way analysis of variance in Table 3 show a significant change in the ratios of LDL/HDL (P=0.00, F=23.986) and VLDL/HDL (P=0.00, F=23.986), as well as the expression of the genes CHOP (P=0.00, F=23.986) and XBP1 (P=0.00, F=23.986). Also, the LSD follow-up test showed a significant decrease in the values of VLDL/HDL, LDL/HDL (in all experimental groups compared to the control group (P = 0.00) and also a significant decrease in the expression of XBP1 genes in the 5-day training groups and the 3 and 5-day training with starvation groups (P = 0.00), but there was no significant change in the starvation group and the 3-day training alone group (P= 0.845 and P = 0.055, respectively). The chaperone gene also showed a significant decrease in all groups except the starvation alone group (P = 0.580) compared to the control group (P = 0.00).

Table 2: Mean and standard deviation of fatty model rats

mean and standard deviation of variables	starvation group + 5 days of training	starvation group + 3days of training	5days training group	3days training group	starvation group	control group
VLDL/HDL	0.286±0.12	0.72±0.31	0.69±0.21	0.94±0.70	0.73±0.21	2.43±0.43
LDL/HDL	0.61±0.28	1.23±0.32	1.03±0.24	1.17±0.69	1.79±0.32	3.69±0.81
СНОР	0.13±0.20	0.22±0.22	0.30±0.23	0.55±0.38	1.08±0.15	1.00±0.00
XBP1	0.06±0.15	0.22±019	0.35±0.31	0.77±0.21	0.97±0.43	1.00±0.00

Figure 1: A B

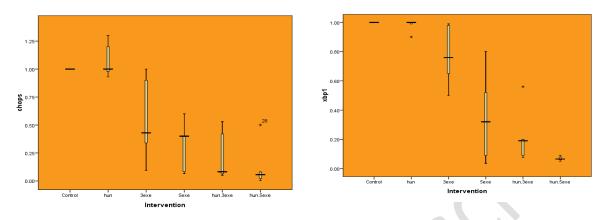


Table 3: One-way analysis of variance test results to compare the means between groups

Variables	Sources of change	sum of squares	mean square	df	f	р
LDL/HDL	Intergroup	140.084	60.39	5	23.986	.000
Mg/dl	within group	6.403	0.252	24		
	total sum	36.240		29		
VLDL/HDL	Intergroup	14.084	2.817	5	19.269	
Mg/dl	within group	3.509	.146	24		.000
	total sum	17.593		29		
	Intergroup	4.103	.821	5	15.340	
СНОР	within group	1.284	.053	24		.000
Ng/mol	total sum	5.386		29		
	Intergroup	4.063	.813	5	26.290	
XBP1	within group	0.742	.031	24		.000
Ng/mol	total sum	4.805	60.39	29		

#### **Discussion**

The aim of the study was the effect of aerobic exercise running on a treadmill and starvation for 4 weeks on the regulation of the apoptotic pathway activity caused by endoplasmic reticulum stress in the liver of male Wistar non-alcoholic fatty model rats. The results of XBP1 description of this research showed that the highest average was observed in the starvation group and the lowest average was observed in the (fat + starvation + 5 days of training) group. The results of the research showed that performing 4 weeks of aerobic exercise along with starvation reduces the expression of both genes involved in the development of NAFLD. While the control and starvation groups have the highest average expression of these genes, exercise decreases the expression. A review of research shows that exercise controls the transcription of XBP1 in the liver (35,36) . Various mechanisms can be involved in this, such as previous studies showing that the goal of starting the UPR is to restore homeostasis and normal ER function. And adaptive mechanisms that increase the expression of genes involved in increasing the capacity to eat ER protein (35). When the primary stimuli that cause UPR not to be eaten are long or excessive, UPR adaptive mechanisms fail. And cell death occurs through apoptosis (37).

It has been reported in different studies that starvation causes a decrease in nutrients inside the cell and it's sensing by brain material-sensing signaling pathways such as mTOR and AMPK pathways, which ultimately stimulates autophagy (39). In addition, p-eiF2α selectively promotes the translation of an increasing number of mRNAs, including ATF4. He does. Activation of IRE1 causes modification of XBP1 and subsequent transcription of molecular chaperones and genesinvolved in ERAD (40). Finally, activated ATF6 undergoes proteolytic cleavage in the Golgi, allowing its mature form to enter the nucleus and ER stress-related genes. Such as ER chaperones and foldases (41). The findings show that starvation activates the IRE1α-XBP1 signal (42). The findings from previous studies show that the combination of fasting diet, acute resistance training and Protein consumption (immediately or 1 hour after exercise stimulation) increases the serum levels of leucine, insulin and glucose, as well as the content of autophagic protein in skeletal muscles (43,44) but it reduces other proteins related to the autophagic pathway in the liver (45). It has also been shown that 6 weeks of wheel running suppressed XBP1s mRNA increase in HFDfed mice, and similar results were shown in mice after 6 weeks of treadmill training (45). In addition, swimming exercise decreased IRE-1a and XBP1 protein levels and decreased hepatic TG content in rats with NAFLD. (47) Lu et al showed that exercise decreased SREBP-1 induced fat accumulation. In the liver through the AMPK pathway to inhibit the mammalian target of rapamycin complex 1 and relieve ERS (46). Overall, exercise reduced hepatic lipogenesis via the PERK/ATF4/SREBP pathway (44,46). These studies show that exercise regulates hepatic XBP1 and SREBPs through ERS signaling and thus reduces fat accumulation in the liver of NAFLD. Exercise helps reduce excessive aberrant phosphorylation in the endoplasmic reticulum that causes apoptosis and cell death. Also, endurance activity such as swimming causes that due to the compatibility between these transmembrane proteins, the amount of misfolded or over folded proteins in the endoplasmic reticulum, which cause stress, decreases. As a result, the amount of stress in the endoplasmic reticulum decreases. Of course, the results of some studies are not consistent with the present study. Among other things, in a research where short-term sports activity such as a one-day sprint or a five-day activity for a week has no effect on the expression of XBP1, ATF6, and PERK proteins, which is probably due to the duration of the sports activity

(43, 45). Because the duration of their activity is less than a week. If it has been shown to express these proteins, the minimum time of sports training should be four weeks. It has also been reported that rats that had a history of sports activity had less stress symptoms and the expression of UPR (XBP1, ATF6) and CHOP genes after resistance training than rats that did not have any sports activity, which shows that Sports activity has a positive effect on reducing stress symptoms in exercise rats (49). Since sports activity is a therapeutic solution to reduce liver diseases including NAFLD. The results of the present study showed that NAFLD increases the expression of XBP1 and CHOP genes, and this increase was significant in the control group compared to the exercise and exercise + starvation groups, which can be a possible confirmation that endoplasmic reticulum stress is one of the One of the main causes of cell apoptosis in the liver. According to the statistical results of our research, it showed a significant decrease in the ratio of lipoproteins (VLDL/HDL, LDL/HDL) in all groups compared to the control group. In recent years, several clinical trials have shown that starvation is an effective way to reduce fat and regulate lipid profile. Starvation or energy-restricted diets have favorable effects on body weight, total fat mass, and liver fat reduction. In addition, intermittent fasting can improve biomarkers of systemic inflammation and appetite-regulating hormones (50). A recent finding suggests that exercise is better than a calorie restriction program in cholesterol biosynthesis. Short-term exercise combined with dietary interventions has a great effect on reducing metabolic risks and fasting insulin levels (50). In a study by Askari et al. in 2012, they showed that 8 weeks of aerobic exercise reduced the percentage of subcutaneous fat, total cholesterol, RF and plasma low-density lipoproteins in non-athletic women (51) Also, in another study by Dadban et al. in 2021, it was shown that 4 weeks of regular aerobic exercise reduced liver lipase and thus reduced triglyceride production in VLD L-C and LDL-C. Elevated LDL-C is an independent risk factor for coronary artery disease, while lowering LDL-C to 60 mg/dL reduces the risk of coronary heart disease by 50% within two years. HDL-C transports cholesterol from peripheral tissues to the liver and then directs excess cholesterol to the bile for excretion (52). However, our results are consistent with the findings of the aforementioned studies. In summary, in such research, understanding the effect of diet on disease severity is one of the most complex aspects in the management of patients with nonalcoholic fatty liver disease. And as a result, evaluating the effect of dietary interventions is challenging because it affects the entire metabolism and it is difficult to isolate specific (beneficial) effects on the liver. However, a low-calorie, low-carbohydrate diet combined with continuous aerobic exercise with repetitions of at least 3 days and up to 5 days of exercise could potentially be suitable for the successful "treatment" of NAFLD.

#### Conclusion

Impaired autophagy may be a critical mechanism in the pathogenesis of nonalcoholic fatty liver disease, and the role of exercise and starvation as an important tool in the prevention of nonalcoholic fatty liver disease. The findings of the research showed that the breakdown of lipids in the liver moderated the disease due to the increase in non-selective autophagy of the liver endoplasmic reticulum. Running with starvation helps NAFLD rats to control the level of CHOP and XBP1 genes and help apoptosis and removal of waste cells as well as reduce ER stress, so this method can be used as a safe and healthy interventional treatment in the treatment of the disease. Liver diseases (NASH, NAFLD) should be considered.

## Acknowledgement

We would like to thank all the colleagues of Golestan University of Medical Sciences who helped us in the laboratory work of this research.

#### **Ethical statement**

This study was conducted using the research ethics code previously approved by Golestan University of Medical Sciences with the ID. IR.GOUMS.REC.1401.005.

## **Conflicts of interest**

There is no conflict and interest by the authors.

## **Funding sources**

It is private and personal financial resources.

## References

- Kalra A, Yetiskul E, Wehrle CJ, et al. Physiology, Liver. [Updated 2023 May 1]. In: StatPearls [Internet].
   Treasure Island (FL): StatPearls Publishing; 2023 Jan-. Available from: https://www.ncbi.nlm.nih.gov/books/NBK535438/
- 2. Heeren J, Scheja L. Metabolic-associated fatty liver disease and lipoprotein metabolism. Mol Metab. 2021 Aug;50:101238. doi: 10.1016/j.molmet.2021.101238. Epub 2021 Apr 20. PMID: 33892169; PMCID: PMC8324684.
- 3. Todosenko N, Khaziakhmatova O, Malashchenko V, Yurova K, Bograya M, Beletskaya M, Vulf M, Gazatova N, Litvinova L. Mitochondrial Dysfunction Associated with mtDNA in Metabolic Syndrome and Obesity. Int J Mol Sci. 2023 Jul 27;24(15):12012. doi: 10.3390/ijms241512012. PMID: 37569389; PMCID: PMC10418437.
- 4. Ge X, Zheng L, Wang M, Du Y, Jiang J. Prevalence trends in non-alcoholic fatty liver disease at the global, regional and national levels, 1990-2017: a population-based observational study. BMJ Open. 2020 Aug 3;10(8):e036663. doi: 10.1136/bmjopen-2019-036663. PMID: 32747349; PMCID: PMC7402189.
- 5. Chen YY, Yeh MM. Non-alcoholic fatty liver disease: A review with clinical and pathological correlation. J Formos Med Assoc. 2021 Jan;120(1 Pt 1):68-77. doi: 10.1016/j.jfma.2020.07.006. Epub 2020 Jul 9. PMID: 32654868
- Caussy C, Aubin A, Loomba R. The Relationship Between Type 2 Diabetes, NAFLD, and Cardiovascular Risk. Curr Diab Rep. 2021 Mar 19;21(5):15. doi: 10.1007/s11892-021-01383-7. PMID: 33742318; PMCID: PMC8805985.
- 7. Fang YL, Chen H, Wang CL, Liang L. Pathogenesis of non-alcoholic fatty liver disease in children and adolescence: From "two hit theory" to "multiple hit model". World J Gastroenterol. 2018 Jul 21;24(27):2974-2983. doi: 10.3748/wjg.v24.i27.2974. PMID: 30038464; PMCID: PMC6054950.
- 8. Zhou L, Shen H, Li X, Wang H. Endoplasmic reticulum stress in innate immune cells a significant contribution to non-alcoholic fatty liver disease. Front Immunol. 2022 Jul 22;13:951406. doi: 10.3389/fimmu.2022.951406. PMID: 35958574; PMCID: PMC9361020.
- 9. Sandes JM, de Figueiredo RCBQ. The endoplasmic reticulum of trypanosomatids: An unrevealed road for chemotherapy. Front Cell Infect Microbiol. 2022 Nov 10;12:1057774. doi: 10.3389/fcimb.2022.1057774. PMID: 36439218; PMCID: PMC9684732.
- 10. Hu H, Tian M, Ding C, Yu S. The C/EBP Homologous Protein (CHOP) Transcription Factor Functions in Endoplasmic Reticulum Stress-Induced Apoptosis and Microbial Infection. Front Immunol. 2019 Jan 4;9:3083. doi: 10.3389/fimmu.2018.03083. PMID: 30662442; PMCID: PMC6328441.
- 11. Choi JA, Song CH. Insights Into the Role of Endoplasmic Reticulum Stress in Infectious Diseases. Front Immunol. 2020 Jan 31;10:3147. doi: 10.3389/fimmu.2019.03147. PMID: 32082307; PMCID: PMC7005066.

- 12. Bhattarai, K.R., Riaz, T.A., Kim, HR. et al. The aftermath of the interplay between the endoplasmic reticulum stress response and redox signaling. Exp Mol Med 53, 151–167 (2021). https://doi.org/10.1038/s12276-021-00560-8
- 13. Fu, X., Cui, J., Meng, X., Jiang, P., Zheng, Q., Zhao, W., & Chen, X. (2021). Endoplasmic reticulum stress, cell death and tumor: Association between endoplasmic reticulum stress and the apoptosis pathway in tumors (Review). Oncology Reports, 45, 801-808. https://doi.org/10.3892/or.2021.7933
- 14. Wu, D., Huang, LF., Chen, XC. et al. Research progress on endoplasmic reticulum homeostasis in kidney diseases. Cell Death Dis 14, 473 (2023). https://doi.org/10.1038/s41419-023-05905-x
- 15. Nakada EM, Sun R, Fujii U, Martin JG. The Impact of Endoplasmic Reticulum-Associated Protein Modifications, Folding and Degradation on Lung Structure and Function. Front Physiol. 2021 May 25;12:665622. doi: 10.3389/fphys.2021.665622. PMID: 34122136; PMCID: PMC8188853.
- 16. Ajoolabady A, Kaplowitz N, Lebeaupin C, Kroemer G, Kaufman RJ, Malhi H, Ren J. Endoplasmic reticulum stress in liver diseases. Hepatology. 2023 Feb 1;77(2):619-639. doi: 10.1002/hep.32562. Epub 2022 May 24. PMID: 35524448; PMCID: PMC9637239.
- 17. Hou W, Nsengimana B, Yan C, Nashan B, Han S. Involvement of endoplasmic reticulum stress in rifampicin-induced liver injury. Front Pharmacol. 2022 Oct 20;13:1022809. doi: 10.3389/fphar.2022.1022809. PMID: 36339603; PMCID: PMC9630567.
- 18. Rada, P., González-Rodríguez, Á., García-Monzón, C. et al. Understanding lipotoxicity in NAFLD pathogenesis: is CD36 a key driver?. Cell Death Dis 11, 802 (2020). https://doi.org/10.1038/s41419-020-03003-w
- 19. Zhao, J., Hu, Y. & Peng, J. Targeting programmed cell death in metabolic dysfunction-associated fatty liver disease (MAFLD): a promising new therapy. Cell Mol Biol Lett 26, 17 (2021). https://doi.org/10.1186/s11658-021-00254-z
- 20. Wei J, Fang D. Endoplasmic Reticulum Stress Signaling and the Pathogenesis of Hepatocarcinoma. Int J Mol Sci. 2021 Feb 11;22(4):1799. doi: 10.3390/ijms22041799. PMID: 33670323; PMCID: PMC7918477.
- 21. Zhao, Q., Zhang, H., Wu, J., Lv, X., Jin, X., & Hu, J. (2020). Melatonin inhibits the endoplasmic reticulum stress-induced, C/EBP homologous protein-mediated pathway in acute pancreatitis. Molecular Medicine Reports, 22, 1647-1655. https://doi.org/10.3892/mmr.2020.11219
- 22. Song MJ, Malhi H. The unfolded protein response and hepatic lipid metabolism in non alcoholic fatty liver disease. Pharmacol Ther. 2019 Nov;203:107401. doi: 10.1016/j.pharmthera.2019.107401. Epub 2019 Aug 13. PMID: 31419516; PMCID: PMC6848795.
- 23. Houghton D., Thoma C., Hallsworth K., et al. Exercise reduces liver lipids and visceral adiposity in patients with nonalcoholic steatohepatitis in a randomized controlled trial. Clinical Gastroenterology and Hepatology. 2017;15(1):96–102.e3.
- 24. Huabin L., Pin S., Yin C. Research on the intervention of NAFLD by baduanjin. Journal of Chengdu university of physical education. 2018;44:79–83.
- 25. Bacchi E., Negri C., Targher G., et al. Both resistance training and aerobic training reduce hepatic fat content in type 2 diabetic subjects with nonalcoholic fatty liver disease (the RAED2 Randomized Trial) Hepatology (Baltimore, Md.) 2013; 58(4):1287-1295.
- 26. Wong V. W. S., Wong G. L. H., Chan R. S. M., et al. Beneficial effects of lifestyle intervention in non-obese patients with non- alcoholic fatty liver disease. Journal of Hepatology. 2018;69(6):1349–1356.
- 27. Jia Y, Yao Y, Zhuo L, Chen X, Yan C, Ji Y, Tao J, Zhu Y. Aerobic Physical Exercise as a Non-medical Intervention for Brain Dysfunction: State of the Art and Beyond. Front Neurol. 2022 May 13;13:862078. doi: 10.3389/fneur.2022.862078. PMID: 35645958; PMCID: PMC9136296.
- 28. Botrus G, Miller RM, Uson Junior PLS, Kannan G, Han H, Von Hoff DD. Increasing Stress to Induce Apoptosis in Pancreatic Cancer via the Unfolded Protein Response (UPR). Int J Mol Sci. 2022 Dec 29;24(1):577. doi: 10.3390/ijms24010577. PMID: 36614019; PMCID: PMC9820188.
- 29. Islam MA, Adachi S, Shiiba Y, Takeda KI, Haga S, Yonekura S. Effects of starvation-induced negative energy balance on endoplasmic reticulum stress in the liver of cows. Anim Biosci. 2022 Jan;35(1):22-28. doi: 10.5713/ab.21.0140. Epub 2021 Jun 23. PMID: 34237916; PMCID: PMC8738926.
- 30. Mansour SZ, Moustafa EM, Moawed FSM. Modulation of endoplasmic reticulum stress via sulforaphane-mediated AMPK upregulation against nonalcoholic fatty liver disease in rats. Cell Stress Chaperones. 2022

- Sep;27(5):499-511. doi: 10.1007/s12192-022-01286-w. Epub 2022 Jul 2. PMID: 35779187; PMCID: PMC9485504.
- 31. Park SM, Kang TI, So JS. Roles of XBP1s in Transcriptional Regulation of Target Genes. Biomedicines. 2021 Jul 8;9(7):791. doi: 10.3390/biomedicines9070791. PMID: 34356855; PMCID: PMC8301375.
- 32. Hetz C, Zhang K, Kaufman RJ. Mechanisms, regulation and functions of the unfolded protein response. Nat Rev Mol Cell Biol. 2020 Aug;21(8):421-438. doi: 10.1038/s41580-020-0250-z. Epub 2020 May 26. PMID: 32457508; PMCID: PMC8867924.
- 33. Cubillos-Ruiz JR, Silberman PC, Rutkowski MR, Chopra S, Perales-Puchalt A, Song M, Zhang S, Bettigole SE, Gupta D, Holcomb K, Ellenson LH, Caputo T, Lee AH, Conejo-Garcia JR, Glimcher LH. ER Stress Sensor XBP1 Controls Anti-tumor Immunity by Disrupting Dendritic Cell Homeostasis. Cell. 2015 Jun 18;161(7):1527-38. doi: 10.1016/j.cell.2015.05.025. Epub 2015 Jun 11. PMID: 26073941; PMCID: PMC4580135.
- 34. Zhao P, Huang P, Xu T, Xiang X, Sun Y, Liu J, Yan C, Wang L, Gao J, Cui S, Wang X, Zhan L, Song H, Liu J, Song W, Liu Y. Fat body Ire1 regulates lipid homeostasis through the Xbp1s-FoxO axis in Drosophila. iScience. 2021 Jul 7;24(8):102819. doi: 10.1016/j.isci.2021.102819. PMID: 34381963; PMCID: PMC8333185.
- 35. Petrescu M, Vlaicu SI, Ciumărnean L, Milaciu MV, Mărginean C, Florea M, Vesa ŞC, Popa M. Chronic Inflammation-A Link between Nonalcoholic Fatty Liver Disease (NAFLD) and Dysfunctional Adipose Tissue. Medicina (Kaunas). 2022 May 6;58(5):641. doi: 10.3390/medicina58050641. PMID: 35630058; PMCID: PMC9147364.
- 36. Heinle JW, DiJoseph K, Sabag A, Oh S, Kimball SR, Keating S, Stine JG. Exercise Is Medicine for Nonalcoholic Fatty Liver Disease: Exploration of Putative Mechanisms. Nutrients. 2023 May 24;15(11):2452. doi: 10.3390/nu15112452. PMID: 37299416; PMCID: PMC10255270.
- 37. Bartoszewska, S., Collawn, J.F. Unfolded protein response (UPR) integrated signaling networks determine cell fate during hypoxia. Cell Mol Biol Lett 25, 18 (2020). https://doi.org/10.1186/s11658-020-00212-1
- 38. Alex S, Boss A, Heerschap A, Kersten S. Exercise training improves liver steatosis in mice. Nutrition & metabolism. 2015 Dec;12(1):1-1. [DOI] [PMID] [PMCID] [Google Scholar]
- 39. Jaud M, Philippe C, Di Bella D, Tang W, Pyronnet S, Laurell H, Mazzolini L, Rouault-Pierre K, Touriol C. Translational Regulations in Response to Endoplasmic Reticulum Stress in Cancers. Cells. 2020 Feb 26;9(3):540. doi: 10.3390/cells9030540. PMID: 32111004; PMCID: PMC7140484.
- 40. Park SM, Kang TI, So JS. Roles of XBP1s in Transcriptional Regulation of Target Genes. Biomedicines. 2021 Jul 8;9(7):791. doi: 10.3390/biomedicines9070791. PMID: 34356855; PMCID: PMC8301375.
- 41. Sharma RB, Snyder JT, Alonso LC. Atf6α impacts cell number by influencing survival, death and proliferation. Mol Metab. 2019 Sep;27S(Suppl):S69-S80. doi: 10.1016/j.molmet.2019.06.005. PMID: 31500833; PMCID: PMC6768497.
- 42. Zhao P, Huang P, Xu T, Xiang X, Sun Y, Liu J, Yan C, Wang L, Gao J, Cui S, Wang X, Zhan L, Song H, Liu J, Song W, Liu Y. Fat body Ire1 regulates lipid homeostasis through the Xbp1s-FoxO axis in Drosophila. iScience. 2021 Jul 7;24(8):102819. doi: 10.1016/j.isci.2021.102819. PMID: 34381963; PMCID: PMC8333185.
- 43. Pinto AP, Vieira TS, Marafon BB, Batitucci G, Cabrera EMB, da Rocha AL, Kohama EB, Rodrigues KCC, de Moura LP, Pauli JR, Cintra DE, Ropelle ER, de Freitas EC, da Silva ASR. The Combination of Fasting, Acute Resistance Exercise, and Protein Ingestion Led to Different Responses of Autophagy Markers in Gastrocnemius and Liver Samples. Nutrients. 2020 Feb 28;12(3):641. doi: 10.3390/nu12030641. PMID: 32121154; PMCID: PMC7146592.
- 44. Debnath, J., Gammoh, N. & Ryan, K.M. Autophagy and autophagy-related pathways in cancer. Nat Rev Mol Cell Biol 24, 560–575 (2023). https://doi.org/10.1038/s41580-023-00585-z
- 45. Zou Y, Qi Z. Understanding the Role of Exercise in Nonalcoholic Fatty Liver Disease: ERS-Linked Molecular Pathways. Mediators Inflamm. 2020 Jul 25;2020:6412916. doi: 10.1155/2020/6412916. PMID: 32774148; PMCID: PMC7397409.
- 46. Lu X, Ding Y, Liu H, Sun M, Chen C, Yang Y, Wang H. The Role of Hydrogen Sulfide Regulation of Autophagy in Liver Disorders. Int J Mol Sci. 2022 Apr 6;23(7):4035. doi: 10.3390/ijms23074035. PMID: 35409395; PMCID: PMC8999478.

- 47. Zhang Y, Liu Y, Liu X, Yuan X, Xiang M, Liu J, Zhang L, Zhu S, Lu J, Tang Q, Cheng S. Exercise and Metformin Intervention Prevents Lipotoxicity-Induced Hepatocyte Apoptosis by Alleviating Oxidative and ER Stress and Activating the AMPK/Nrf2/HO-1 Signaling Pathway in db/db Mice. Oxid Med Cell Longev. 2022 Sep 9;2022:2297268. doi: 10.1155/2022/2297268. PMID: 36120597; PMCID: PMC9481363.
- 48. Romero-Gómez, M.; Zelber-Sagi, S.; Trenell, M. Treatment of NAFLD with Diet, Physical Activity and Exercise. J. Hepatol. 2017,67, 829–846.
- 49. Cheang WS, Wong WT, Zhao L, Xu J, Wang L, Lau CW, Chen ZY, Ma RC, Xu A, Wang N, Tian XY, Huang Y. PPARδ Is Required for Exercise to Attenuate Endoplasmic Reticulum Stress and Endothelial Dysfunction in Diabetic Mice. Diabetes. 2017 Feb;66(2):519-528. doi: 10.2337/db15-1657. Epub 2016 Nov 17. PMID: 27856609.
- 50. Thomas Marjot ,Jeremy W Tomlinson, Leanne Hodson, David W Ray: Timing of energy intake and the therapeutic potential of intermittent fasting and time-restricted eating in NAFLD. 2023;72:1607–1619.doi:10.1136/gutjnl-2023-329998 https://gut.bmj.com/content/gutjnl/72/8/1607.full.pdf.
- 51. Askari A, Askari B, Fallah Z, Kazemi Sh . Effect of eight weeks aerobic training on serum lipid and lipoprotein levels in women, Journal of Gorgan University of Medical Sciences / 32 Spring 2012 / vol 14 / no 1.
- 52. Minoo Dadban Shahamat , Asra Askari , Ramezan Arab Koohsar ;effects of Four Weeks of High-Intensity Intermittent Training and Continuous Walking on Atherogenic Indices of Obese Middle-Aged Men, mljgoums ,Volume 15, Issue 2 ,(Mar-Apr 2021, 15(2): 42-47.