Inhibitory Effect of Probiotic Bacteria Isolated from Camel Milk on Clinical Strains of Drug-Resistant Helicobacter pylori

ABSTRACT

Background and Objectives: Helicobacter pylori is the most common cause of gastritis and ulcer worldwide. Treatment of such infections may lead to failure due to drug resistance. This study aimed to investigate the antimicrobial effects of bacteria present in camel milk on the growth of drug-resistant clinical isolates of H. pylori.

Methods: In this cross-sectional study, biopsy samples from 75 patients with digestive symptoms were transferred to laboratory in transport medium containing homogeneous compounds. In order to isolate H. pylori, urease-positive biopsies were promptly cultured in brucella agar enriched with defibrinated sheep blood and fetal calf serum. Disk diffusion agar test was used to evaluate antibiotic susceptibility and agar well diffusion method was applied to study the antagonistic effect of probiotics isolated from camel milk on the H. pylori isolates.

Results: The frequency of H. pylori isolates was 42.7%. The highest rate of resistance was observed against metronidazole (56.3%). In addition, the rate of resistant to amoxicillin, ciprofloxacin, and clarithromycin and tetracycline was 31.3%, 18.8%, 15.6%, respectively. Lactobacillus plantarum (59.3%) was more frequent than other Lactobacillus species. L. plantarum, Lactobacillus fermentum and Lactobacillus casei showed favorable inhibitory effects against the H. pylori isolates, but L. plantarum (with inhibition zone diameter of 20.3 mm) showed the highest inhibitory effect.

Conclusion: Considering the increasing rate of drug resistance and the inhibitory effect of probiotics isolated from milk, health providers recommend that promoting consumption of probiotic food seems beneficial for the general population and those suffering from gastrointestinal disorders.

Keywords: Helicobacter pylori, Drug resistance, Camel, milk, Probiotics.

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INTRODUCTION

Today, *Helicobacter pylori* is one of the most common causes of gastric infections and ulcers in the world, and 40-50% of humans have been affected by this bacterium. According to the World Health Organization, *H. pylori* is recognized as a type 1 carcinogen (1-3). The bacterium is abundant in the gastric mucosa of carriers and can continue its growth by penetrating into the mucin layer. It can also infect gastric mucosa for decades in spite of acquired immune responses, inflammation and the continuous alternation of gastric epithelial cells (2).

*H. pylori* is a curved, gram-negative, motile and non-spore-forming bacterium with 4-6 polar flagella. The bacterium grows under microaerophilic conditions. High humidity and temperature of 35-37 °C are the optimal growth conditions. Most strains require 3-5 days and sometimes 7 days of incubation (1). Since the bacteria are located under the protecting mucin layer, only antibiotics that could reach beneath the mucous membrane are effective (4). Amoxicillin, tetracycline, metronidazole and clarithromycin along with proton pump inhibitors and bismuth salts are usually used for treatment of *H. pylori* infections. However, the side effects and antibiotic resistance may sometimes lead to treatment failure.

Regarding the risk of drug resistance in various infections, it is recommended using a combination of various antimicrobial compounds to overcome this issue. Among these compounds, probiotics are clinically effective and economically justified alternatives. Besides, they lack the side effects of common antibiotics. Probiotics are live microorganisms with positive effects on microecology of the digestive system and help maintain human health (5). Beneficial role of probiotics in treatment of gastrointestinal diseases such as acute diarrhea has been confirmed (6). Probiotics exert these beneficial effects through various mechanisms such as non-immunologic mechanisms (antimicrobial substances, competing for adherence to target cells and help to reinforce the mucosal barrier), immunologic mechanisms and secretion (bacteriocins). In general, probiotics create pores in the phospholipid bilayer by affecting the cytoplasmic membrane and generating proton momentum (7). Some of the final products of lactic fermentation such as lactic acid, acetic acid and hydrogen peroxide have antibacterial effects. Production of great amounts of lactate may also have inhibitory effects on pathogenic bacteria (8).

Camel milk is a rich source of probiotics, such as lactobacilli. It has been used to treat respiratory diseases such as tuberculosis and asthma, as well as gastrointestinal and digestive disorders. In some African countries (Egypt, Sudan, Kenya and Somali), cameleers believe that drinking camel milk makes humans stronger and faster (7). Camel milk is also rich in amino acids including valine, methionine, lysine, arginine and phenylalanine. It also contains high amounts of albumin and globulin (3).

The aim of this study is to investigate the in vitro antagonistic effects of probiotics isolated from camel milk on the growth of clinical strains of *H. pylori* resistant to selective agents.

MATERIAL AND METHODS

In this cross-sectional study, we collected endoscopic specimens from 75 patients with stomach pain, duodenal ulcers, gastritis and the suspected symptoms of gastric cancer referred to two gastroenterology centers in Tehran during 2016-2017. After taking biopsy specimens from gastric antrum of each patient, the specimens were transferred to laboratory in a broth glycolate transport medium (Merck, Germany) containing homogeneous compounds. The samples were screened by initial tests such as rapid urease test and complementary tests. Urease-positive biopsy samples were cultured on brucella blood agar (Merck, Germany) containing polymyxin, vancomycin and trimethoprim, and then incubated in microaerophilic conditions (10% CO₂) at 37 °C for 5-6 days. The medium was enriched with 7% fetal calf serum and defibrinated sheep blood. Next, the suspected *H. pylori* colonies grown on the medium were confirmed based on morphology, Gram staining and positive urease, catalase, and oxidase reaction.

Antibiotic susceptibility testing was performed by the agar disk diffusion method (Kirby-Bauer method) using metronidazole (5 µg), amoxicillin (30 µg), ciprofloxacin (5 µg), tetracycline (10 µg) and clarithromycin (2 µg) disks purchased from Mast Co., UK. In this test, 100 µl of bacterial suspension equal to 3
McFarland standard of *H. pylori* resistant to antibiotics was prepared, and then spread on brucella blood agar (Merck, Germany). Then, wells (about 6 mm in diameter) were created in the medium using sterile Pasteur pipette and 100 μl of the upper bacterial suspension isolated from milk was poured into the wells. The plates were incubated at 37 °C for 48 hours under microaerophilic conditions. Next, the diameter of inhibition zone was measured and recorded in millimeters. Each step of the tests was repeated three times. In addition, *Leuconostoc mesenteroides* PTCC1663 standard strain was used for comparison.

Chi-Square and paired t-test were used in SPSS (version 20) for data analysis. Statistical significance level was set at 0.05.

Susceptibility tests were examined by error categorization.

**RESULTS**

*H. pylori* isolates were obtained from adolescents (age range: 20-59 years, mean age: 35±8.2 years, Male/Female: 1:1.7). Of the gastric biopsy specimens taken from patients, 32 (42.7%) cases were found to be *H. pylori*. These cases were from patients with gastritis (75%), peptic ulcers (19%) and gastric cancer (6%) (Figure 1).

The *H. pylori* isolates were most susceptible to amoxicillin and clarithromycin and most resistant against metronidazole (Table 1).

The camel milk samples were collected in sterile conditions from 3- and 7-year-old camels. The samples were transferred to laboratory in sterile containers. In order to isolate probiotic bacteria, the milk samples were inoculated into MRS broth (Merck, Germany) and M17 broth (Merck, Germany). After incubation in anaerobic conditions at 37 °C for 48 hours, the solutions enriched in the MRS and M17 agar were cultured and the microscopic and macroscopic properties of isolates were examined. Species were identified based on the pattern of glucose fermentation and the colonies of the isolated lactic acid bacteria were inoculated in special broth. Paraffin was added to the medium and the tube was incubated at 37 °C for 4 days (the time required for production of antibacterial material). Later, paraffin was extracted and the contents of the tubes were transferred to sterile glass tubes. The tubes were centrifuged at 3500 RPM for 15 minutes and the supernatant was preserved for further studies.

Agar well diffusion method was used to study antimicrobial effects of probiotics. For this purpose, bacterial suspension equal to 1 McFarland standard of *H. pylori* resistant to antibiotics was prepared, and then spread on brucella blood agar (Merck, Germany). Then, wells (about 6 mm in diameter) were created in the medium using sterile Pasteur pipette and 100 μl of the upper bacterial suspension isolated from milk was poured into the wells. The plates were incubated at 37 °C for 48 hours under microaerophilic conditions. Next, the diameter of inhibition zone was measured and recorded in millimeters. Each step of the tests was repeated three times. In addition, *Leuconostoc mesenteroides* PTCC1663 standard strain was used for comparison.

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The majority (96.3%) of lactic acid isolates belonged to *Lactobacillus* species. Among these, *Lactobacillus plantarum* was the most frequent isolate (59.3%) and the other strains were *Lactobacillus fermentum* and *Lactobacillus casei* (Table 2).

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**Figure 1- Prevalence of gastrointestinal complications in patients**

*Gastritis* 75%, *Ulcer* 19%, *Cancer* 6%
The antagonistic effect of bacteriocins generated by lactobacilli was evaluated after 3-5 days of incubation. Measurement of the inhibition zone indicated the complete susceptibility of drug-resistant *H. pylori* isolates to lactobacilli in laboratory conditions (Table 3).

Inhibition zone was formed upon introduction of *L. plantarum*, *L. fermentum* and *L. casei* to the growing antibiotic-resistant *H. pylori*. The average diameter of inhabitation zone was 21.3, 19.21 and 17.2 mm for *L. plantarum*, *L. fermentum* and *L. casei*, respectively, which confirmed the inhibitory effect of these probiotics on the resistant strains of *H. pylori* (Figure 2). However, Lactococci had not inhibitory effects on the resistant *H. pylori* strains.

**DISCUSSION**

*H. pylori* is regarded as one of the most common causes of gastrointestinal infections worldwide. About 40-50% of the world’s population is affected by this bacterium (8). *H. pylori* can also cause gastric diseases such as chronic gastritis, peptic ulcer, gastric cancer and lymphoma of mucosa-associated lymphoid tissue (10). In the present study, 42.7% of the strains were identified as *H. pylori*. This indicates the high resistance of *H. pylori* strains to most commonly used antibiotics. In our study, 18.8% of the strains were resistant to ciprofloxacin, which is less than the rates reported by studies in Germany (11), Portugal (12), France (13) and Spain (14). Given that ciprofloxacin is used as the second-line therapy for treatment of *H. pylori*, the high resistance rate observed against this antibiotic...
In a study on the antagonistic effect of L. casei on H. pylori and salmonella, it was revealed that L. casei inhibited the expression of H. pylori flagellin genes A and B, which ultimately prevented from bacterial movement and resulted in morphology change to coccoid form (22). In the present study, most lactobacilli isolated from the milk of dromedary were related to L. plantarum, L. fermentum and L. casei.

In a study conducted in Morocco, 37.7% and 25.8% of strains isolated from the dromedary milk were lactobacilli and lactococci respectively (23). We found that the H. pylori isolates resistant to all five antibiotics were susceptible to bacteriocins of lactobacilli isolated from camel milk. Similar to our study, Heidari et al. confirmed the antibacterial effect of bacteriocin produced by Lactobacillus isolates from diaries on 10 pathogenic bacteria using well diffusion assay (24). Another study has also confirmed the inhibitory effect of bacteriocin produced by lactobacilli isolates from milk on H. pylori (25). Khonafari et al. studied the production of Lactocins by probiotic strains in local yogurt samples. The results of this study showed the isolation of 21 strains of lactic acid bacteria. Production of antimicrobial combinations by strains in the logarithmic phase of their growth and their antimicrobial effects were also observed (26). Mojgani et al. confirmed the inhibitory properties of bacteriocin produced by L. acidophilus isolates from cheese samples in Karaj (Iran) against some pathogenic bacteria (27). In another study, leuconostocs, lactobacilli and enterococci were the most frequent lactic acid bacteria present in milk of Iranian dromedary (28). However, we found L. plantarum as the most frequent species. A study in 2015 investigated the in vitro antagonistic effects of L. casei on antibiotic-resistant H. pylori and confirmed the susceptibility of this pathogen to Lactobacillus (29).

Given the similarity with our findings, it should be noted that we isolated lactobacilli strains from camel milk and investigated the species resistant to multiple antibiotics.

CONCLUSION
Based on the results, it can be concluded that the rate of resistance to selective antimicrobial agents used for treatment of gastritis is increasing. Given the
high resistance of H. pylori isolates to antibacterial agents and the inhibitory effects of lactic acid bacteria, the addition or presence of lactobacilli in fermented foods such as camel milk could have protective effects against infections that are caused by H. pylori. Thus, it is recommended to promote the consumption of such products to help improve public health.

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CONFLICT OF INTEREST
The authors declare no conflict of interest.


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